Short Reports 3543

organic layer was subjected to CC on silica gel eluting with CHCl₃. Fractions containing 1 were subjected to repeated preparative TLC using cyclohexane–CH₂Cl₂ (1:4) to afford yellow needles of 1 (5 mg, 0.00006%), mp 205–206°; UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 215 (4.42), 230sh (4.16), 254 (4.38), 344 (3.66) and 370sh (3.63) nm; UV $\lambda_{\max}^{\text{MeOH}}$ nm (log ε): 204 (4.68), 225 (4.37), 240sh (4.18), 272 (4.29), 344 (3.51) and 435 (3.75); IR ν_{\max}^{KBr} cm⁻¹: 3270, 1750, 1640, 1505 and 1485; ¹H NMR (CDCl₃, 400 MHz), (TMS): δ 7.90 (d, J = 8.2 Hz, 1H, H-7), 7.38 (d, J = 8.2 Hz, 1H, H-6), 5.96 (s, 1H, OH, D₂O exch, C-12 OH), 3.47 (sept, J = 6.9 Hz, 1H, H-15), 2.79 (s, 3H, H-20) and 1.34 (d, J = 6.9 Hz, 6H, H-16 and H-17); MS (electron impact, 70 eV) m/z (rel. int.): 242 [M]⁺ (96), 227 (100), 213 (27), 199 (9), 184 (6), 171 (12), 171 (12), 155 (9), 141 (21), 129 (38) and 115 (30); HRMS 242.0938 for C₁₅H₁₄O₃, cal. 242.0932.

Acknowledgements—The authors would like to thank our colleagues in the Department of Analytical Chemistry of the Shanghai Institute of Materia Medica, for IR, UV, NMR and

MS measurements, and the Division of Cancer Treatment, National Cancer Institute, Bethesda, MD, for financial support.

REFERENCES

- Lin, L.-Z., Wang, X.-M., Huang, X.-L., Huang, Y. and Yang, B.-J. (1988) Acta Pharm, Sinica 23, 273.
- Lin, L.-Z., Wang, X.-M., Huang, X.-L., Huang, Y. and Yang, B.-J. (1988) Planta Med. 54, 443.
- 3. Lin, L.-Z., Wang, X.-M., Huang, X.-L. and Huang, Y. (1989) *Acta Chimica Sinica* (in press).
- Blaskó, G., Lin, L.-Z. and Cordell, G. A. (1989) J. Org. Chem. 53, 6113.
- Lin, L.-Z., Blaskó, G. and Cordell, G. A. (1989) Phytochemistry 28, 177.
- Lin, L.-Z. and Cordell, G. A. (1989) Phytochemistry 28 (in press).
- Karanatsios, D., Scarpa, J. S. and Eugster, C. H. (1966) Helv. Chim. Acta, 49, 1151.

Phytochemistry, Vol. 28, No. 12, pp. 3543–3544, 1989. Printed in Great Britain,

0031 9422/89 \$3.00 + 0.00 © 1989 Pergamon Press plc

PHLOROACYLPHENONES IN THE ESSENTIAL OIL OF THRYPTOMENE SAXICOLA

KIM A. DASTLIK, EMILIO L. GHISALBERTI and PHILLIP R. JEFFERIES

Department of Organic Chemistry, The University of Western Australia, Nedlands, 6009, Western Australia

(Received in revised form 12 May 1989)

Key Word Index—Thryptomene saxicola; Myrtaceae; essential oil; phloroacylphenones; O-isobaeckeol.

Abstract—The essential oil of *Thryptomene saxicola* is shown to contain α -pinene, 1,8-cineole, α -terpineol, globulol, *O*-isobaeckeol and its homologue.

INTRODUCTION

The genus *Thryptomene* is endemic to Australia and consists of ca 25 species [1] most of which are restricted to the south-west and Eremean provinces of Western Australia [2]. Previous work on Thryptomene had been restricted to the detection and isolation of sesquiterpenes. Thus the essential oil of *T. kochi* has been shown to contain aromadendrene and globulol [3, 4]. The presence of the latter in *T. australis* Endl and *T. stenocalyx* F.v.M. has also been established [4]. As part of a screening programme of essential oils from Western Australian plants, we had the opportunity of analysing the essential oil of a sample of *T. saxicola* (A. Cunn.) Schau which from preliminary examination appeared to contain two aromatic compounds. These have been characterized as *O*-isobaeckeol (1) and its homologue (2).

RESULTS AND DISCUSSION

Steam distillation of the leaves and terminal branches of the plant yielded a yellow oil (1.5%) which on distillation afforded fractions of α -pinene (57%), a mixture (12%) of α -pinene, cineole, α -terpineol and five unidentified sesquiterpene hydrocarbons. A third fraction (20%) contained three compounds. Separation by chromatography on basic alumina gave globulol and another compound to which the structure of O-isobaeckcol (1) was assigned on the following evidence.

Interpretation of the spectral data indicated that the compound ($[M]^+$ 238) contained a pentasubstituted benzene with two methoxyls, a methyl, a hydrogen bonded hydroxyl (δ_H 13.14) and an isobutanoyl group as substituents. These features are consistent with the compound being backeol (3). However, comparison of their

3544 Short Reports

Table 1. Carbon chemical shifts of baeckeol (3) and O-iso-baeckeol (1)

C	3	1	C	3	1
1	210.54	210.75	4'	163.07°	164.10
2	39.60	38.40	5'	85.79	107.51 ^b
3	19.29	19.58	6'	163.99°	164.06
2-Me	19.29	19.58	Ar-Me	7.24*	8.50
1′	105,85a	107.51 ^b	OMe	55.38	62.02
2'	160.89	160.14	OMe	55.38	55.83
3'	104.86a	95.76			

a-cValues with identical superscript may be interchanged.

¹³C NMR spectra (Table 1) and ¹H NMR spectra showed significant differences. In particular the carbon of the aromatic methyl in 3 at δ_C 7.24 appears at lower field (δ_C 8.50) in 1 as does the unsubstituted aromatic carbon (δ_C 85.79 in 3; δ_C 95.76 in 1). This suggests the orientation of substitutents for the compounded to be that shown in 1 which is named O-isobaeckeol. This compound has been prepared [5] but has not been found to occur naturally before.

Crude fractions of 1 were shown by GC-MS to contain trace amounts of another compound at longer R_t . The mass spectrum showed an [M]⁺ at m/z 252, otherwise it was indistinguishable from that of 1, suggesting it to be a homologue probably with an isopentanoyl side chain (2). The presence of 2 is not unexpected given the occurrence of leptospermone (4) and its homologue, flavesone in species of the Myrtaceae. It has also been suggested that the homologue of baeckeol might be a natural product [6].

EXPERIMENTAL

General experimental details have been described elsewhere [7].

Isolation of O-isobaeckeol (1) from Thryptomene saxicola. Leaves and terminal branches (750 g) of the air-dried plant, obtained from Australian Flower Farms, Coorow, Western Australia, were steam distilled for 7 hr to give a yellow oil (12 ml). GC analysis (HP Ultra-1 column, cross-linked methyl silicon gum, 0.02 mm × 50 m WCOT) of the crude essential oil showed the following composition: α -pinene (60%), 1,8-cineole (7.5%), α terpineol (6%), five unidentified sesquiterpene hydrocarbons (11%), globulol (6%), O-isobaeckeol (3.5%) and its homologue (trace). Distillation (<140°C, 15 mm) yielded a colourless oil (3.4 g) whose ¹H NMR and mass spectra were identical with those of authentic α -pinene. The residue (2.5 g) was separated into three fractions by Kügelrohr distillation (250°, 15 mm) which by GC-MS were shown to contain the following: Fraction 1 (150 mg) α -pinene, 1,8-cineole, α -terpineol and five sesquiterpene hydrocarbons ([M] + m/z 204); fraction 2 (528 mg) contained all the components of the undistilled sample; fraction 3

3

(411 mg)—mainly globulol, O-isobaeckeol (1) and the homologue (2). GC-MS (X-linked Me-silicone gum, $0.31 \text{ mm} \times 25 \text{ m}$ WCOT capillary 100-250° at 4°/min) of fraction 3 showed two peaks at $R_t > 20$ min. The major one at R_t 22 min was shown to be 1 and the minor one at R_t 23.9 min gave the following MS m/z(rel. int.) 252 [M] + (9), 195 (100), 180 (3), 152 (7), 151 (2), 109 (2), 69 (2). Chromatography of fraction 3 on basic alumina (act. I; 10 g) and elution with 10% CH₂Cl₂: petrol yielded 1 (100 mg) as an oil, R_t 0.6 (CH₂Cl₂; silica gel), v_{max} (CHCl₃) 1625–1610 cm⁻¹; ¹H NMR (300 MHz, CDCl₃) δ 13.14 (1H, br s, OH), 6.25 (1H, s, H-3'), 3.84 (3H, s, OCH₃), 3.82 (1H, septet, J = 6.8 Hz, H-2), 3.71 (3H, s, OMe), 2.06 (3H, s, 5'-Me), 1.18 (6H, d, J = 6.7 Hz, 2-Me and H₃-3); ¹³C NMR (75.5 MHz, CDCl₃): see Table 1. MS (EI), m/z (rel. int.): 238 [M] + (11), 195 (100), 180 (3), 152 (9), 151 (2), 69 (2), 43 (2). Elution with CH₂Cl₂ afforded fractions of a single compound (150 mg), R_f 0.2 (CH₂Cl₂, silica gel) whose ¹H, ¹³C NMR, mass spectra and GC R, were identical with those of an authentic sample of globulol.

REFERENCES

- Willis, J. C. (1973) A Dictionary of the Flowering Plants and Ferns 8th Edn (revised by H. K. Airy-Shaw). Cambridge University Press, Cambridge.
- 2. Burbidge, N. T. (1963) Dictionary of Australian Plant Genera, Angus and Robertson, Sydney.
- Graham, A., Jefferies, P. R., Melrose, G. J. H., Thieberg, K. J. L. and White, D. E. (1960) Aust. J. Chem. 13, 372.
- Melrose, G. J. H. (1960) Ph.D. Thesis. The University of Western Australia.
- Schiemenz, G. P., Beherens, H., Ebert, C. P., Maienschein, K. and Schroder, J.-M. (1985) Z. Naturforsch 40b, 681.
- Schiemenz, G. P. and Schmidt, U. (1976) Liebigs Ann. Chem. 1514.
- Ghisalberti, E. L., Jefferies, P. R., Mori, T. A., Skelton, B. W. and White, A. H. (1985) Tetrahedron 41, 2517.

^{*}Assigned by selective decoupling of methyl protons.